This article was downloaded by: On: *25 January 2011* Access details: *Access Details: Free Access* Publisher *Taylor & Francis* Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Sulfur Chemistry

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713926081

The Toxicity of Sulfonium Compounds

P. Kestell^a; S. C. Mitchell^b

^a Cancer Research Laboratory, Auckland Medical School, University of Auckland, Auckland, New Zealand ^b Department of Pharmacology and Toxicology, St. Mary's Hospital Medical School, Imperial College of Science, Technology and Medicine, University of London, London, Great Britain

To cite this Article Kestell, P. and Mitchell, S. C.(1996) 'The Toxicity of Sulfonium Compounds', Journal of Sulfur Chemistry, 17: 2, 159 – 176

To link to this Article: DOI: 10.1080/01961779608047891 URL: http://dx.doi.org/10.1080/01961779608047891

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

Sulfur Reports. Volume 17, pp. 159–181 Reprints available directly from the publisher Photocopying permitted by license only © 1996 OPA (Overseas Publishers Association) Amsterdam B.V. Published in The Netherlands by Harwood Academic Publishers GmbH Printed in Malaysia

THE TOXICITY OF SULFONIUM COMPOUNDS

P. KESTELL' and S. C. MITCHELL²

¹Cancer Research Laboratory, Auckland Medical School, University of Auckland, Private Bag 92019, Auckland, New Zealand and ²Department of Pharmacology and Toxicology, St. Mary's Hospital Medical School, Imperial College of Science, Technology and Medicine, University of London, Norfolk Place, London W2 1PG, Great Britain

(Received June 7, 1995)

Sulfonium compounds and their related salts are one class of charged sulfur compounds which play important rôles within biology. Those naturally occurring sulfonium compounds which are essential for biochemical processes within living systems have been summarised. Synthetic compounds, some of which are analogues of established pharmacological agents, possess a variety of biological activities and have been investigated for many potential therapeutic applications including antitumour and neurological properties. Recent developments within this field have been reviewed together with the adverse toxicological implications arising from the biological generation of reactive intermediates and subsequent enzyme interaction.

Key words: S-Adenosyl-L-methionine, dimethyl-β-propiothetin, S-methyl-L-methionine, sulfocholine, sulfonium compounds, toxicity

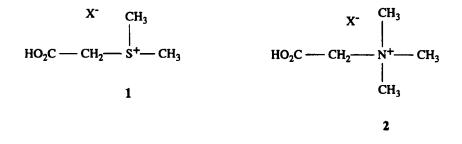
TABLE OF CONTENTS

1.	INT	RODUCTION	160
2.	STI	RUCTURAL CHARACTERISTICS AND CHEMISTRY	160
3.	SO	URCES OF SULFONIUM COMPOUNDS	163
	3.1.	Naturally Occurring Compounds	163
		3.1.1. S-Adenosyl-L-methionine [S-(5'-deoxyadenosyl-(5')-L-methionine]	163
		3.1.2. S-Methyl-L-methionine (methionine methylsulfonium)	164
		3.1.3. Dimethyl-β-propiothetin (2-carboxyethyldimethylsulfonium)	164
		3.1.4. Sulfocholine (2-hydroxyethyldimethylsulfonium salt) derivatives	165
	3.2.	Synthetic Sulfonium Compounds	165
		3.2.1. Analogues of known pharmaceutical compounds	165
			166
		3.2.3. Analogues of neuroactive compounds	167
4.	то	KICOLOGICAL INTERACTIONS	168
	4.1.	Enzyme Interaction	168
	4.2.	Formation of Reactive Intermediates	170
	4.3.	Sulfonium Compounds in the Metabolism Process	172

REFERENCES	174
SUBJECT INDEX	177
AUTHOR INDEX	179

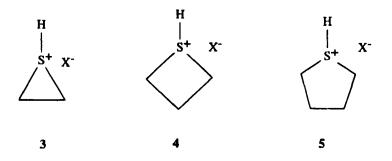
1. INTRODUCTION

The first synthetic sulfonium compounds prepared over a century ago^{1,2} possessed quite simple chemical structures based on (carboxymethyl)dimethylsulfonium salts 1, making them comparable to betaine, [(carboxymethyl)trimethylammonium hydroxide (inner salt)] 2, one of the quaternary ammonium compounds found in plants. Knowledge of the remarkable biological properties, both pharmacological and toxicological, of these charged ammonium derivatives undoubtedly provided the incentive for the synthesis and examination of analogous sulfur compounds.^{3,4} Astutely aware of this relationship, the researchers called their new class of sulfur-containing compounds thetines, as they explained, "We have given the substances the name thetine to recall its relation to betaine and the fact that it contains sulphur".^{1,2}



2. STRUCTURAL CHARACTERISTICS AND CHEMISTRY

The parent compound of this group is the simple sulfonium ion H_3S^+ which is structurally analogous to the hydroxonium (hydronium; oxonium) ion H_3O^+ . Substitution of all the hydrogen atoms with organic groups such as methyl or ethyl results in the formation of the respective sulfonium derivatives, trimethylsulfonium and triethylsulfonium. The sulfur atom in all of these compounds is formally trivalent bearing an overall positive charge, and the outer electronic cover is best considered as having five electrons which reside within three bonds and one lone pair. Most simple sulfonium compounds are pyramidal, but introduction of bulky substituents (eg. phenyl) distorts the conformation as a consequence of steric repulsion.⁵ Sulfonium compounds may also exist as cyclic structures, the simplest being the three-membered ring associated with the thiiranium or episulfonium ion **3**. Other ring systems which have been identified and studied include the four-membered thietanium ion **4** and the five-membered thiolanium ion **5**.

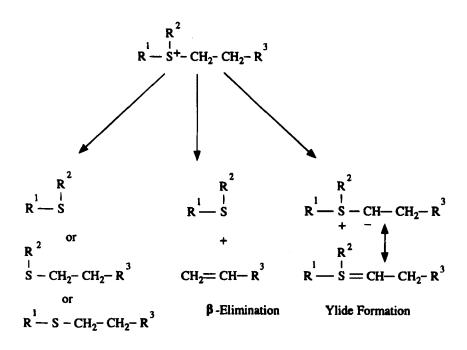


The sulfonium compounds were the first by which the potential optical activity of sulfur derivatives was established. Optical resolution of the camphorsulfonate and bromocamphorsulfonate of ethylmethylthetine^{6,7} into their enantiomeric forms at the turn of the century were landmark discoveries in sulfur chemistry. It was realised that these enantiomers only had three groups attached to the active centre, the forth being a true ion, the sulfonium pole. The discovery of optically active sulfoxides^{8,9} made it certain that a tricovalent sulfur atom could form an asymmetric molecule, raising the importance of the lone pair of electrons. A large number of optically pure sulfonium salts have been prepared, either by resolution or by stereospecific synthesis.¹⁰

The chemical properties of sulfonium salts are well understood and have been more than adequately reviewed in the literature.¹¹⁻¹⁵ The most important reactions applicable to the biological field are (*i*) nucleophilic displacement in which there is C-S⁺ bond fission, (*ii*) β -elimination which yields an olefin and a thioether, and (*iii*) α -deprotonisation resulting in ylide formation in which the sulfur atom stabilises the adjacent carbanion¹⁶ (Scheme 1). One of the most important features of sulfonium salts is their relative instability in solution, particularly in alkaline environments. Complex mixtures of decomposition products are formed, presumably as a result of the sulfonium salt undergoing one or more of the above mentioned reactions.

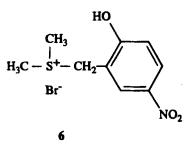
This in turn has contributed to numerous difficulties associated with the isolation, characterisation and quantitation of these compounds. By their very nature sulfonium salts are polar, water-soluble, ionic compounds; factors which do not assist their analysis. A variety of techniques have been employed with limited success such as gravimetric analysis, titrimetric and colorimetric methods, thin-layer and paper chromatography, polarography and ultraviolet spectroscopy.¹⁷ However, recent advances in high-pressure liquid chromatography have made possible the separation and quantitation of sulfonium compounds with diverse chemical structures in different types of biological and chemical matrices. Of particular importance has been the development and application of ion chromatography using conductivity detection¹⁸⁻²¹ as well as conventional reverse phase or cation-exchange chromatography employing ultraviolet detection.²²⁻²⁶

The chemical reactivity of the sulfonium group has been found in certain instances to be advantageous when specifically incorporated into a derivatising reagent. Such compounds have found numerous applications not only in analytical chemistry but also in protein and



Nucleophilic Displacement

Scheme 1

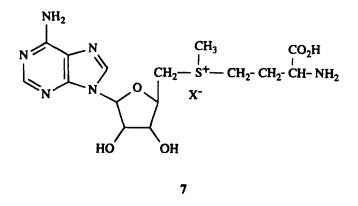


enzyme biochemistry. One such reagent, dimethyl (2-hydroxy-5-nitrobenzyl)sulfonium bromide 6 was found to react selectively with tryptophanyl residues in proteins under mild conditions.²⁷ This compound has proved to be a useful tool by which the role of this amino acid moiety can be studied in relation to the functional activity of an enzyme. Some enzyme systems which have been studied in this way include human plasminogen,^{28,29} human thrombin,³⁰ human antithrombin,³¹⁻³³ rabbit muscle creatine kinase,³⁴ rabbit skeletal muscle myosin subfragment (S-1),^{35,36} bovine pancreatic carboxypeptidase A,³⁷ yeast lactate dehydrogenase³⁸ and yeast glyceraldehyde-3-phosphate dehydrogenase.³⁹ The strong alkylating properties of trimethylsulfonium and triethylsulfonium hydroxide have been used to derivatise theophylline,⁴⁰ pentachloro- and tetrachlorophenols⁴¹ as well as bacterial fatty acids^{42,43} prior to their analysis by gas-liquid chromatography.

3. SOURCES OF SULFONIUM COMPOUNDS

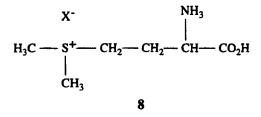
3.1. Naturally Occurring Compounds

3.1.1. S-Adenosyl-L-methionine (S-(5'-deoxyadenosyl-(5')-L-methionine) S-Adenosyl-L-methionine 7 and its demethylated derivative [S-adenosyl-(5')-3-methylthiopropylamine] appear to be widely distributed throughout nature, virtually all living tissues contain these compounds.⁴⁴ Within intermediary metabolism S-adenosyl-L-methionine is involved in so many of the cellular metabolic reactions that its function and status in cellular terms has been compared to that of adenosine triphosphate. It is a compound for which new roles are being discovered continually and the subject of many excellent books and reviews.⁴⁴⁻⁵¹



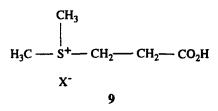
S-Adenosyl-L-methionine is known to interact with the catecholamine (β -adrenergic, dopaminergic) and related serotonin (5-hydroxytryptamine) neurotransmitter pathways within the central nervous system, leading to elevated serotonin and 5-hydroxyindoleacetic acid levels in some regions of the rat brain.⁵⁰ The continual search for compounds which alter catecholamine and indoleamine metabolism in the brain, deviations in which have been proposed as a biochemical basis for depression, led S-adenosyl-L-methionine to undergo clinical evaluation as an antidepressant agent⁵² where it was found to be equally effective in treating the disorder when compared to standard regimens employing tricyclic antidepressants. S-Adenosyl-L-methionine has also been proffered to treat malnourished patients, especially with liver disease and cholestasis⁵³ as well as having anti-inflammatory and analgesic activities.^{49,50}

3.1.2. S-Methyl-L-methionine (methionine methylsulfonium) This compound 8 has been found exclusively within the plant kingdom, being isolated from many vegetables such as cabbage, kohlrabi (turnip-cabbage) and related crops where it may account for up to 0.2% of the tissue dry weight⁵⁴ as well as fruit,⁵⁵ potato⁵⁶ and green tea.⁵⁷ Evidence suggests that it plays a significant role as a methyl donor during the biosynthesis of methionine and that it may also serve as a source of ethylene during the ripening of fruit and the development of flowers. Its presence in cow's milk probably results via direct transferance from the herbivore's diet.⁵⁸



S- Methyl-L-methionine was originally established as having anti-ulcer properties when extracts of cabbage leaves were shown to prevent the development of experimentally induced gastric ulcers in guinea-pigs. The active factor extracted from fresh cabbage juice was termed vitamin U (anti-gizzard erosion factor; ulcer-preventative factor) and for a time it was fashionable to drink cabbage-water in the hope of enouraging the healing of peptic ulcers. Attempts to establish the effectiveness of S-methyl-L-methionine in the treatment of ulcers in man on a rational clinical basis has led to many studies⁵⁹⁻⁶² especially. in Russia and Japan where other related clinical applications such as the stimulation of skin regeneration and wound healing⁶³ and anti-inflammatory properties are also being investigated.^{64,65}

3.1.3. Dimethyl- β -propiothetin (2-carboxyethyldimethylsulfonium) Dimethyl- β -propiothetin 9 appears to be restricted to aquatic life and has been extracted from many different forms of algae, plankton and marine invertebrates.⁶⁶⁻⁶⁹ The fact that these minute creatures are the principal foods of many fish and filter-feeders has presumably led the detection of dimethyl- β -propiothetin in molluscs, crustacea and fish.^{70,71} It was the first sulfonium compound to be isolated in pure form from a living organism; the marine algae *Polysiphonia fastigiata* and *Polysiphonia nigrescens*.^{68,72} A related tertiary sulfonium compound, 4-dimethylsulfonio-2-methoxybutanoate, has also been identified in marine algae.⁷³ Both of these compounds, when present, occur in very high concentrations, dimethyl- β -propiothetin comprising up to 2–9% of the wet weight of some algae,^{66,74} suggesting that they have important biological functions, such as overcoming salt or water stress within these organisms.⁷⁵



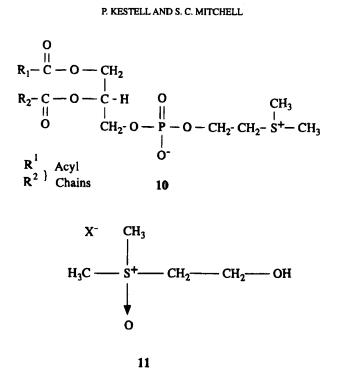
Owing to structural similarities with S-methyl-L-methionine, dimethyl- β -propiothetin has also been examined as an anti-ulcer agent and results suggest that it is more effective than the latter compound.^{76,77} The methyl ester of dimethyl- β -propiothetin also stimulates the parasympathetic nervous system.

3.1.4. Sulfocholine (2-hydroxyethydimethylsulfonium salt) derivatives The novel sulfolipid, phosphatidyl sulfocholine **10**, has been found in a variety of marine diatoms, Cylindrotheca fusiformis, Navicula pelliculosa, Nitzschia alba, Phaeodactylum tricornutum⁷⁸⁻⁸⁰ and appears to replace the function of the ubiquitous phospholipid, phosphatidyl choline, in cell membranes.⁸¹ The absence of nitrogen-containing phospholipids (eg. phosphatidyl choline, phosphatidyl ethanolamine, phosphatidyl serine) encountered in other living organisms suggests an evolved intermediary metabolic system where the charged sulfur moiety replaces the more usual quaternary ammonium arrangement. The sulfocholine fragment probably arises from methionine via dimethyl- β -propiothetin, which, as previously mentioned, is present in many marine organisms.⁸² Perhaps these single-celled algae (Bacillariophyta) have retained a closer link to the common sulfur-metabolising progenitor of life; an interesting curiosity.

2-Hydroxyethydimethylsulfoxonium salt 11, which is the sulfoxide of sulfocholine, occurs in the marine bryozoan *Alcyonidium gelatinosum*. These small creatures live in aggregate colonies but maintain separate identities, each having an identifiable alimentary canal. The sulfonium salt is thought to act as a hapten; on repeat exposure it initiates a hypersensitivity reaction and is responsible for causing a form of eczema and contact dermatitis (colloquially known as the 'Dogger Bank itch') which is prevalent amongst North Sea fishermen.⁸³³⁴

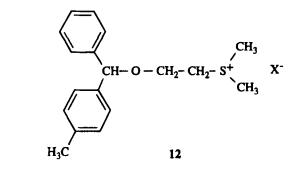
3.2. Synthetic Sulfonium Compounds

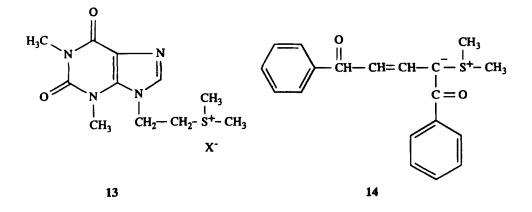
3.2.1. Analogues of known pharmaceutical compounds Exploring the mechanisms of action of pharmacological agents amidst the search for more potent derivatives with lower incidences of side-effects, has led to the development of several sulfonium analogues of current therapeutic agents, where the sulfur moiety becomes the important heteroatom at the functional site.



Sulfonium analogues of diphenhydramine hydrochloride, a widely employed antihistamine used for the symptomatic relief of allergy, also exhibited antihistamine activity with one compound 12 being five times more effective than the parent nitrogen-containing molecule.^{85,86} A sulfonium analogue of the ophylline, the iodide salt of 7-(β -methylthioethyl)theophylline 13, has been shown to have the same but weaker pharmacological properties as theophylline,⁸⁷ and sulfonium derivatives of 1-aroyl-4-oxo-4-aryl-2-butenylides 14 posses antimicrobial activity.⁸⁸ The sulfonium 15 and 2-benzylsulfonium 16 analogues of platelet activating factor (PAF) display a high degree of cytotoxicity towards HL60 cells whilst exhibiting weak hypotensive and platelet aggregating activity.89

3.2.2. Antitumour activities The structure-activity relationships and drug design of dimethylsulfonium compounds as biological response modifiers and as antitumour agents is a subject of contemporary interest. In particular, tris(2-chloroethyl)sulfonium halides and alkylbis(2-chloroethyl)sulfonium halides, which have highly reactive alkylating functional groups, exhibited a growth suppressing effect on two tumour cell lines, the Yoshida ascites sarcoma of rats and the Ehrlich carcinoma in mice.^{90,91} A recently developed dimethylsulfonium compound, 'Suplatast Tosilate®' 17, designed to treat allergy disorders, acts as a biological response modifier and suppresses IgE synthesis in both mice and humans.⁹²⁻⁹⁴ Other sulfonium compounds reported to have antitumour activity in experimental systems include a number of substituted phenylacyl sulfonium bromides⁹⁵ and the monovalent

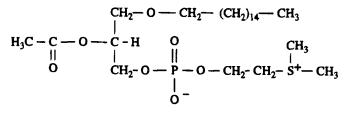


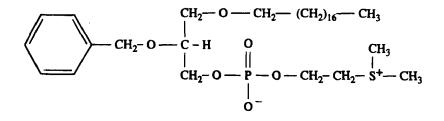


lipophilic cation 2,6-bis(4-aminophenyl)-4-[4-(dimethylamino)phenyl]thiopyrylium chloride 18.⁶⁶ Clinical activity appears to be restricted to the major component of the antitumour drug 'Bleomycin', bleomycin A_2 19, which has within its complex molecular structure a sulfonium group. This compound is an important clinical agent used to treat head and neck tumours, testicular cancer and Hodgkin's disease.^{97,98}

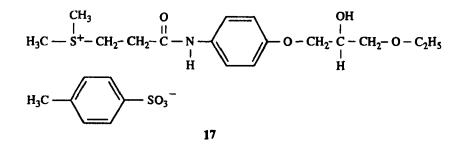
3.2.3. Analogues of neuroactive compounds In neurological studies, where sulfonium derivatives have been examined for their action within neural tissue, it has generally been found that the sulfonium analogues exhibit the same effect as their nitrogen-containing counterparts but to a lesser degree.⁹⁹

This can be exemplified by the sulfonium analogue of choline, sulfocholine, which can be substituted for choline in phospholipid biosynthesis^{81,100} and serve as a substrate for choline acetyltransferase to yield acetylsulfocholine,¹⁰¹ the latter compound possessing pharmacological properties resembling acetylcholine but being less potent.¹⁰² In addition, the sulfonium analogues of polymethylenebis(trialkylammonium) salts and succinylcholine, all neuroactive compounds, have been synthesised and evaluated as neuromuscular blocking agents. Amonst these were the salts of hexamethylene-1,6-bis(dimethylsulfonium) **20**,







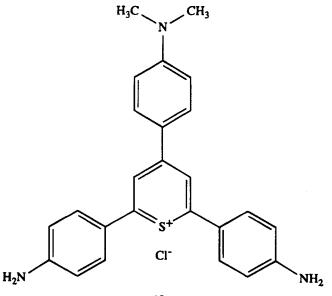


¹⁰³ decamethylene-1,10-(dimethylsulfonium) 21 ¹⁰⁴ and succinylbis(sulfocholine) 22
 ¹⁰⁵ which, like their more potent nitrogen counterparts, were found to possess the ability to block neuromuscular transmission.

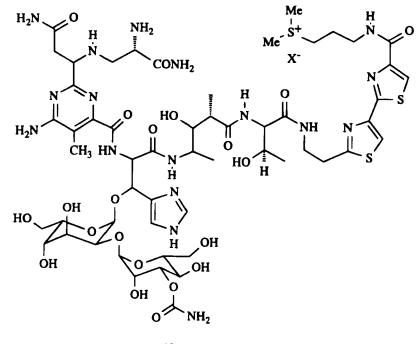
4. TOXICOLOGICAL INTERACTIONS

4.1. Enzyme Interaction

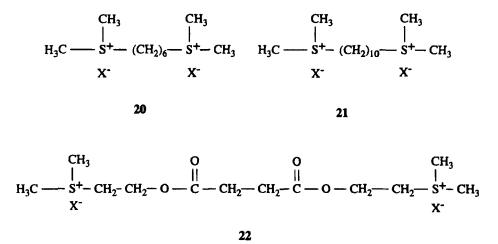
A number of sulfonium salts have been found to be specific cellular enzyme inhibitors. It has been shown that both triphenylsulfonium chloride and the monovalent lipophilic cation 2,6-bis(4-aminophenyl)-4-[4-(dimethylamino)phenyl]thiopyrylium chloride 18 are



18



19



selectively taken up by the mitochondria of cells and there inhibit oxidative phophorylation and adenosine triphosphatase activity as well as the electron-transfer system in the NAD-cytochrome b regions of the respiratory chain.^{96,106} These interactions have been postulated to account for the tumoricidal properties of 2,6-bis(4-aminophenyl)-4-[4-(dimethylamino)phenyl]thiopyrylium chloride.

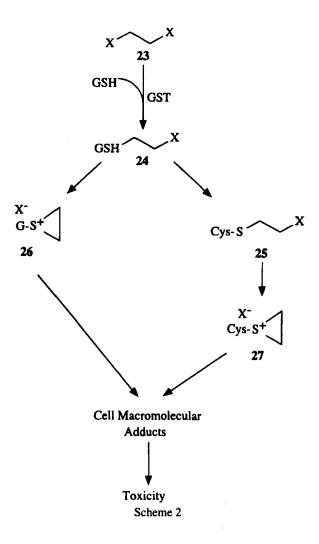
Compounds such as decamethylenebis(dimethylsulfonium) bromide and *n*-octadecyldimethylsulfonium bromide are potent inhibitors of phospholipase C, as are several other alkysulfonium and alkyldisulfonium salts.^{107,10} As mentioned previously, sulfonium compounds are also capable of interfering with neuronal activity. Thus bis- and poly-onium molecules which contain sulfonium moieties, as well as monoalkylsulfonium compounds such as trimethylsulfonium iodide, display anticholinesterase activity as well as neuromuscular blocking activity.^{109,110} Such compounds must be considered as potentially toxic and treated with due care.

4.2. Formation of Reactive Intermediates

Whilst few direct toxicological studies have been undertaken on sulfonium compounds the formation of sulfonium intermediates and their interaction with cellular processes have featured in a number of toxicological mechanisms.

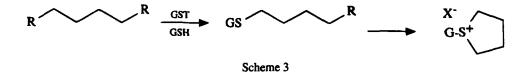
It has been proposed that thiiranium or episulfonium ions are the ultimate toxic species associated with the toxicity of 1,2-dihaloethanes **23**,^{51,111,112} a group of compounds once commonly used as soil fumigants, gasoline additives, solvents and synthetic intermediates. These compounds are now known to be mutagenic and carcinogenic and to be responsible for the production of tumours at multiple sites including the adrenal glands, kidney, liver, lungs, mammary glands, skin and stomach.¹¹³⁻¹¹⁶

The 1,2-dihaloethanes are probably metabolised to the corresponding S-(2-haloethyl)glutathione conjugate 24 via glutathione S-transferase catalysed displacement of a halogen (Scheme 2). As expected, these tripeptide conjugates are degraded to the cysteine conjugate 25 by metabolic pathways associated with mercapturic acid formation. Both the glutathione and cysteine conjugates may be considered as biosynthetic sulfur half-mustards which have strong alkylating and electrophilic properties attributable to neighbouring group interactions in nucleophilic displacement reactions.^{117,118} Consequently, highly reactive episulfonium ions 26,27 are formed through internal displacement of the second halogen atom by the sulfur atom. It is postulated that these intermediate species react with biological macromolecules forming covalent adducts which



in turn give rise to toxicity.^{119,120} Other haloalkyl compounds such as 1,2-dichloropropane¹²¹ and 1,2-dibromo-3-chloropropane¹²² are assumed to elicit toxicity by forming similar reactive episulfonium intermediates.

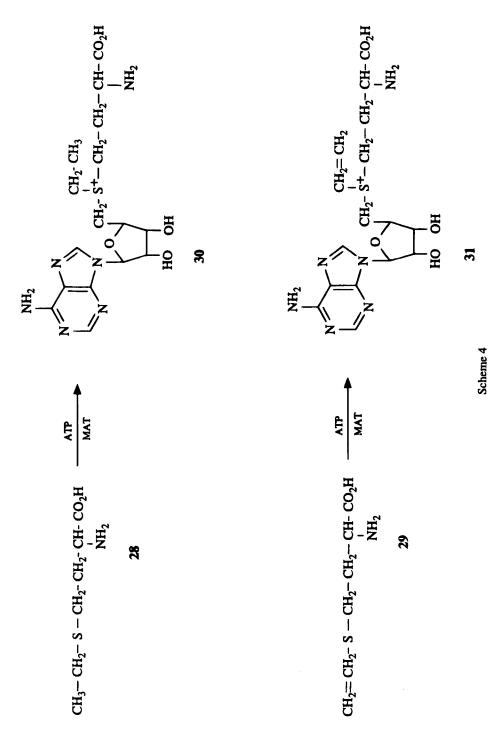
However, compounds such as 1,4-dihaloalkanes, for example the chemotherapeutic agent, 1,4-butanediol dimethanesulfonate¹²³⁻¹²⁷ and the known mutagens 1,4-dibromobutane and 1,4-diiodobutane¹²⁸⁻¹³⁰ are also mainly metabolised by glutathione conjugation, forming an unstable conjugate which rapidly cyclises to a five-membered thiolanium ion glutathione conjugate (Scheme 3). In comparison to the episulfonium ion, the thiolanium ion is chemically a more stable entity and therefore unlikely to react with celular macromolecules. The mechanism of toxicity of these particular compounds must lie elsewhere.



4.3. Sulfonium Compounds in the Metabolism Process

Bioactivation has also been evoked in the hepatocarcinogenicity of ethionine 28 and vinthionine 29. The metabolic formation of their corresponding S-adenosylated derivatives 30 and 31 is thought to lead to mimicking of endogenous S-adenosyl-L-methionine thereby permitting enzymatic transfer of ethyl or vinyl groups to macromolecules such as DNA, with carcinogenic sequelae (Scheme 4). S-Adenosylvinthionine would be expected to be much more reactive towards cellular nucleophiles than either vinthionine or S-adenosylethionine because of the inductive and electron-sharing stabilisation of the carbanion intermediate formed during nucleophilic addition at the β -carbon atom of the vinyl group.^{51,131}

A role for sulfonium intermediates in the metabolism of epoxides has also been proposed. It is suggested that methionine (or other methylthio cellular components) react with the epoxide to form methylsulfonium ions which undergo elimination reactions to yield methylthio metabolites.^{51,132} Other evidence suggests that S-adenosyl-L-methionine may be directly involved in biotransformation. An S-adenosyl-L-methionine dependent thioether transferase has been identified in mice which can metabolise several alkyl sulfides including 2-chloroethyl ethyl sulfide^{133,134} and a non-specific S-adenosyl-L-methionine dependent N-methyltransferase has been identified in rabbit liver which can metabolise a wide range of amines.⁵¹



Downloaded At: 12:37 25 January 2011

173

REFERENCES

- 1. C. Brown and E. A. Letts, Trans. R. Soc. Edinb. 28, 571 (1878).
- 2. E. A. Letts, Trans. R. Soc. Edinb. 28, 618 (1878).
- 3. A. C. Brown and T. R. Fraser, J. Anat. Physiol. 2, 224 (1866-1867).
- 4. A. C. Brown and T. R. Fraser, Trans. R. Soc. Edinb. 25, 151 (1867-1869).
- M. Simonetta and A. Gavezzotti, in: C. J. M. Stirling and S. Patai (Eds.), The Chemistry of the Sulphonium Group, Wiley, Chichester, 1981, pt 1, pp. 1-14.
- 6. W. J. Pope and S. J. Peachey, J. Chem. Soc. 77, 1072 (1900).
- 7. S. Smiles, J. Chem. Soc. 77, 1174 (1900).
- 8. P. W. B. Harrison, J. Kenyon and H. Phillips, J. Chem. Soc. 2079 (1926).
- 9. J. Holloway, J. Kenyon and H. Phillips, J. Chem. Soc. 3000 (1928).
- K. K. Andersen, in: C. J. M. Stirling and S. Patai (Eds.), The Chemistry of the Sulphonium Group, Wiley, Chichester, 1981, pt 1, pp. 229-266.
- V. Du Vigneaud, A Trail of Research in Sulphur Chemistry and Metabolism in Related Fields, Cornell University Press, New York. 1952.
- 12. F. Challenger, Aspects of the Organic Chemistry of Sulphur, Butterworths, London. 1959.
- 13. F. Schlenk, Fortschr. Chem. Organ. Natur. 23, 61 (1965).
- C. J. M. Stirling, in: S. Oae (Ed.), Organic Chemistry of Sulfur, Plenum Press, New York and London, 1977, pp. 473–525.
- 15. C. J. M. Stirling and S. Patai, The Chemistry of the Sulphonium Group, Wiley, Chichester, 1981.
- A. C. Knipe, in: C. J. M. Stirling and S. Patai (Eds.), The Chemistry of the Sulphonium Group, Wiley, Chichester, 1981, pt 1, pp. 313-385.
- M. R. F. Ashworth, in: C. J. M. Stirling and S. Patai (Eds.), The Chemistry of the Sulphonium Group, Wiley, Chichester, 1981, pt 2, pp. 79–99.
- 18. G. Aced, E. Anklam and H. J. Mockel, J. Liq. Chromat. 10, 3321 (1987).
- 19. G. Aced, H. J. Mockel and S. F. Nelson, J. Liq. Chromat. 12, 3201 (1989).
- 20. J. L. Hoffman, J. Chromat. 588, 211 (1991).
- 21. R. J. Williams, J. Chromatogr. Sci. 20, 560 (1982).
- 22. Z. Zappia, M. Carteni-Farina, P. Galletti and F. Della Ragione, Methods Enzymol. 94, 57 (1983).
- 23. J. Gorham, J. Chromat. 287, 345 (1984).
- 24. R. K. Gordon, G. A. Miura, T. Alonso and P. K. Chaing, Methods Enzymol. 143, 191 (1987).
- 25. B. Guattari, J. Chromat. 567, 254 (1991).
- 26. J. Lagendijk, J. B. Ubbink and W. J. H. Vermaak, J. Chromat. 576, 95 (1992).
- 27. H. R. Horton and W. P. Tucker, J. Biol. Chem. 245, 3397 (1970).
- 28. S. M. Hochschwender and R. A. Laursen, J. Biol. Chem. 256, 11172 (1981).
- 29. M. Llinas, A. De Marco, S. M. Hochschwender and R. A. Laursen, Eur. J. Biochem. 135, 379 (1983).
- 30. G. B. Villanueva, Biochemistry 20, 6519 (1981).
- 31. N. M. Blackburn and C. C. Sibley, J. Biol. Chem. 255, 824 (1980).
- 32. G. I. Karp, J. A. Marcum and R. D. Rosenberg, Arch. Biochem. Biophys. 233, 712 (1984).
- 33. M. F. Scully, N. Shah, V. Ellis and V. V. Kakker, Thrombosis Haemostasis, 65, 351 (1991).
- 34. H. M. Zhou and C. L. Tsou, Biochim. Biophys. Acta 830, 59 (1985).
- 35. M. M. Werber, Y. M. Peyser and A. Muhlrad, Biochemistry 26, 2903 (1987).
- Y. M. Peyser, A. Muhlrad and M. M. Werber, FEBS Lett. 259, 346 (1990).
- 37. L. N. Y. Liu-Wu and R. Horton, Biochim. Biophys. Acta 577, 22 (1979).
- U. Mayer, R. Hensel, M. Deparade, H. E. Pauly, G. Pfleiderer and W. E. Trommer, Eur. J. Biochem. 126, 549 (1982).
- 39. H. D. Heilman and G. Pfleiderer, Biochim. Biophys. Acta 384, 331 (1975).
- 40. B. L. Lee, P. Jacob and N. L. Benowitz, J. Chromatog. 494, 109 (1989).
- 41. W. Butte, M. Kirsch and J. Denker, Int. J. Environ. Anal. Chem. 13, 141 (1983).
- 42. K.-D. Muller, H. Husmann and H. P. Nalik, Int. J. Med. Microbiol. 274, 174 (1990).
- 43. H. P. Nalik, K.-D. Muller and R. Ansorg, J. Med. Microbiol. 36, 371 (1992).
- 44. R. J. Huxtable, Biochemistry of Sulfur, Plenum Press, New York, 1986, pp. 75-76.
- F. Salvatore, E. Boreck, V. Zappia, H. G. Williams-Ashman and F. Schlenk, The Biochemistry of Adenosylmethionine, Columbia University Press, New York, 1977.
- 46. E. Usdin, R. T. Borchardt and C. R. Creveling, Transmethylation, Elsevier, New York, 1979.

- 47. E. Usdin, R. T. Borchardt and C. R. Creveling, Biochemistry of S-Adenosylmethionine and Related Compounds, Macmillan, London, 1982.
- G. A. Maw, in: C. J. M. Stirling and S. Patai (Eds.), The Chemistry of the Sulphonium Group, Wiley, Chichester, 1981, pt 2, pp. 703-770.
- 49. G. Stramentinoli, Am. J. Med. 83, 35, (1987).
- G. Stramentinoli, in: R. T. Borchardt, C. R. Creveling and P. M. Ueland (Eds.), Biological Methylation and Drug Design, Humana Press, New Jersey, 1986, pp. 315-326.
- P. A. Crooks, in: L. A. Damani (Ed.), Sulphur-Containing Drugs and Related Organic Compounds: Chemistry, Biochemistry and Toxicology, Ellis Horwood Ltd, Chichester, 1989, Vol. 1, pt. B. pp. 155-180.
- R. K. Chawla, H. L. Bonkovsky and J. T. Galambos, *Drugs*, 40, 98 (1990).
 P. Almasio, M. Bortolini, L. Pagliaro and M. Cortolti, *Drugs*, 40, 111 (1990).
- 54. E. G. Kovacheva, Prikl. Biokhim. Mikrobiol. 10, 129 (1974).
- 55. F. F. Wong and J. F. Carson, J. Agric. Food Chem. 14, 247 (1966).
- 56. G. Werner, R. Hossli and H. Neukom, Lebensm. Wiss. Technol. 2, 145 (1969).
- 57. T. Kiribushi and T. Yamanishi, Agric. Biol. Chem. 27, 56 (1963).
- 58. T. W. Keenan and R. C. Lindsay, J. Dairy Sci. 51, 112 (1968).
- 59. E. Strehler, Gastroenterologia 84, 119 (1955).
- 60. T. Tanaka, J. Ohara, K. Takezoe, T. Ando and N. Hokari, New Remedies Therapies, 8, 20 (1969).
- 61. Y. Yoshinaka and M. Nakamura, Pharmacometrics 21, 921 (1981).
- 62. A. S. Salim, J. Pharm. Pharmac. 39, 553 (1987).
- 63. U. Cucinotta, Boll. Soc. Ital. Biol. Sper. 35, 1142 (1959).
- 64. G. Stramentinoli, C. Pezzoli and E. Catto, Minerva Med. 66, 4434 (1975).
- 65. M. Cortellaro, Minerva Med. 63, 1854 (1972).
- 66. R. G. Ackman, C. S. Tocher and J. McLachlan, J. Fish. Res. Bd. Can. 23, 357 (1966).
- 67. F. Challenger, R. Bywood, P. Thomas and B. J. Hayward, Arch. Biochem. Biophys. 69, 514 (1957).
- 68. F. Challenger and M. L. Simson, J. Chem. Soc. 1591 (1948).
- 69. R. Bywood and F. Challenger, Biochem. J. 53, Abs. 26 (1953).
- 70. R. G. Ackman and H. J. Hingley, J. Fish. Res. Bd. Can. 25, 267 (1968).
- 71. T. Motohiri, Mem. Fac. Fish. Hokkaido Univ. 10, 1 (1962).
- 72. P. Hass, Biochem. J. 29, 1297 (1935).
- 73. S. Sciuto, M. Piattelli and R. Chillemi, Phytochemistry 21, 227 (1982).
- 74. G. L. Cantoni, D. G. Anderson and E. Rosenthal, J. Biol. Chem. 222, 171 (1956).
- 75. G. Blunden and S. M. Gorden, Prog. Phycol. Res. 4, 39 (1986).
- 76. Y. Ishida, Y. Ogihara and S. Okabe, Jap. J. Pharmac. 54, 333 (1990).
- 77. K. Nakajima, J. Nutr. Sci. Vitaminol. 37, 229 (1991).
- 78. R. Anderson, M. Kates and B. E. Volcani, Nature 263, 51 (1976).
- 79. R. Anderson, M. Kates and B. E. Volcani, Biochim. Biophys. Acta 528, 89 (1978).
- P. Bisseret, S. Ito, P. A. Tremblay, B. E. Volcani, D. Dessort and M. Kates, *Biochim. Biophys. Acta* 796, 320 (1984).
- 81. K. S. Bjerve and J. Bremer, Biochim. Biophys. Acta 176, 570 (1969).
- 82. R. Anderson, M. Kates and B. E. Volcani, Biochim. Biophys. Acta 573, 557 (1979).
- 83. J. S. Carle and C. Christophersen, Toxicon 20, 307 (1982).
- 84. J. S. Carle, H. Thybo and C. Christophersen, Contact Dermatitis 8, 43 (1982).
- 85. O. Exner, M. Borovicka and M. Protiva, Coll. Czech. Chem. Commun. 18, 270 (1953).
- 86. M. Protiva and O. Exner, Coll. Czech. Chem. Commun. 16, 689 (1951)
- 87. J. Maj, H. Sowinska and M. Sypniewska, Archiv. Immunol. Ther. Expt. 10, 125 (1962).
- 88. R. G. Reddy, R. G. Reddy and S. Sarada, Biochem. Int. 19, 215 (1989).
- 89. M. Kates, G. A. Adams, M. L. Bank and F. Snyder, Lipids 26, 1095 (1991).
- 90. A. Lüttringhaus, J. Kimmig, H. Machatzke and M. Jänner, Arzneim. Forsch. 9, 748 (1959).
- 91. A. Lüttringhaus and H. Machatzke, Arzneim. Forsch. 13, 366 (1963).
- 92. N. Matsuura, H. Mori, H. Nagai and A. Koda, Fol. Pharmac. Japon. 100, 485 (1992).
- 93. Y. Yanagihara, M. Kiniwa, K. Ikizawa, T. Shida, N. Matsuura and A. Koda, Jpn. J. Pharmacol. 61, 31 (1993).
- Y. Yanagihara, M. Kiniwa, K. Ikizawa, H. Yamaya, T. Shida, N. Matsuura and A. Koda, Jpn. J. Pharmacol. 61, 23 (1993).
- 95. H. A. Rutter, J. Am. Chem. Soc. 5905 (1951).
- 96. X. Sun, J. R. Wong, K. Song, J. Hu, K. D. Garlid and L. B. Chen, Cancer Res. 54, 1465 (1994).
- 97. P. R. Twentyman, Pharmacol. Therap. 23, 417 (1984).

- B. A. Chabner, in: B. A. Chabner and J. M. Collins (Eds.), Cancer Chemotherapy--Principles and Practise JB Lippincott Company, Philadelphia. 1990. pp. 341-355.
- 99. D. Della Bella, Arch. Ital. Sci. Farmacol. 13, 70 (1963).
- 100. G. A. Maw and V. Du Vigneaud, J. Biol. Chem. 176, 1029 (1948).
- 101. L. Frankenberg, D. Heimbürger, C. Nilsson and B. Sörbo, Eur. J. Pharmacol. 23, 37 (1973).
- 102. H. R. Ing, P. Kordic and D. P. H. Tudor Williams, Brit. J. Pharmacol. 7, 103 (1952).
- 103. D. Della Bella, Naunyn-Schmiedebergs Arch. Exp. Pathol. Pharmakol. 226, 335 (1955).
- 104. J. Walker, J. Chem. Soc. 193 (1950).
- D. Della Bella, R. Villani and G. F. Zuanazzi, Naunyn-Schmiedebergs Arch. Exp. Pathol. Pharmakol. 229, 536 (1956).
- 106. R. H. Barrett and M. J. Selwyn, Biochem. J. 156, 315 (1976).
- 107. P. R. Young, W. R. Snyder and R. F. McMahon, Lipids 26, 957 (1991).
- 108. P. R. Young, W. R. Snyder and R. F. McMahon, Biochim. Biophys. Acta 1121, 297 (1992).
- 109. S. M. Kirpekar, J. J. Lewis and T. C. Muir, Biochem. Pharmacol. 11, 937 (1962).
- 110. S. G. Cohn, S. B. Chishti, J. L. Elkind, H. Reese and J. B. Cohen, J. Med. Chem. 28, 1309 (1985).
- 111. P. J. Van Bladderen, I. M. Bruggeman, W. M. F. Jongen, A. G. Scheffer and J. H. Temmink, in: D. J. Benford, J. W. Bridges and G. G. Gibson (Eds.), Drug Metabolism—From Molecules to Man Taylor and Francis, London, 1987, pp. 151-170.
- 112. W. Dekant and S. Vamvakas, Xenobiotica 23, 873 (1993).
- 113. W. A. Olsen, R. T. Haberman, E. K. Weisburger, J. M. Ward and J. H. Weisburger, J. Nat. Can. Inst. 51, 1993 (1972).
- 114. E. K. Weisburger, Environ. Health Persect. 21, 7 (1977).
- 115. R. D. Storer and R. B. Conolly, Carcinogenesis 4, 1491 (1983).
- 116. L. C. K. Wong, J. M. Winston, C. B. Hong and H. Plotnick, Tox. Appl. Pharmacol. 63, 155 (1982).
- 117. D. R. Dohn and J. E. Casida, Bioorgan. Chem. 15, 115 (1987).
- 118. P.A. Jean and D. J. Reed, Chem. Res. Toxicol. 2, 455 (1989).
- P. J. Van Bladderen, D. D. Briemer, G. M. Rotteveel-Smijs, R. A. W. Jong, W. Bujis, A. van der Gen and G. R. Mohn, *Biochem. Pharmacol.* 29, 2975 (1980).
- 120. W. Webb, A. Elfarra, R. Thom and M. W. Anders, Pharmacology 27, 228 (1985).
- 121. M. J. Bartels and C. Timchalk, Xenobiotica 20, 1035 (1990).
- 122. P. G. Pearson, E. J. Soderlund, E. Dybing and S. D. Nelson, Biochemistry 29, 4971 (1990).
- 123. J. J. Roberts and G. P. Warwick, Nature 183, 1509 (1959).
- 124. J. J. Roberts and G. P. Warwick, Biochem. Pharmacol. 6, 217 (1961).
- 125. J. J. Roberts and G. P. Warwick, Biochem. Pharmacol. 6, 205 (1961).
- 126. M. Hassan and H. Ehrsson, Drug Metab. Dispos. 15, 399 (1987).
- 127. M. Hassan and H. Ehrsson, Eur. J. Drug Metab. Pharmacokinet. 12, 71 (1987).
- 128. D. H. Marchand and M. M. Abel-Monem, Biochem. Biophys. Res. Commun. 128, 360 (1985).
- 129. D. H. Marchand, R. P. Remmel and M. M. Abel-Monem, Drug Metab. Dispos. 16, 85 (1988).
- 130. W. Okenhout, W. M. G. M. van Loon, W. Buijs, A. van der Gen and N. P. E. Vermeulen, Drug Metab. Dispos. 14, 608 (1986).
- 131. W. R. Leopold, J. A. Millar and E. C. Millar, Cancer Res. 42, 4364 (1982).
- 132. L. S. Sheng, E. C. Horning and M. G. Horning, Drug. Metab. Dispos. 12, 297 (1984).
- 133. N. M. Mozier and J. L. Hoffman, Fed. Am. Soc. Expt. Biol. J. 4, 3329 (1990).
- 134. N. M. Mozier, K. P. McConnel and J. L. Hoffman, J. Biol. Chem. 263, 4527 (1988).